

Limonoids from the Leaves and Twigs of *Walsura yunnanensis*Kai-Long Ji,<sup>†,‡</sup> Ping Zhang,<sup>†</sup> Hua-Bin Hu,<sup>†</sup> Shuai Hua,<sup>†</sup> Shang-Gao Liao,<sup>\*,§</sup> and You-Kai Xu<sup>\*,†</sup><sup>†</sup>Key Laboratory of Tropical Plant Resource and Sustainable Use, Xishuangbanna Tropical Botanical Garden, Chinese Academy of Sciences, Menglun, Mengla, Yunnan 666303, People's Republic of China<sup>‡</sup>University of Chinese Academy of Sciences, Beijing 100049, People's Republic of China<sup>§</sup>Engineering Research Center for the Development and Application of Ethnic Medicines and TCM, School of Pharmacy, Guiyang Medical College, Guiyang, Guizhou 550004, People's Republic of China

## S Supporting Information

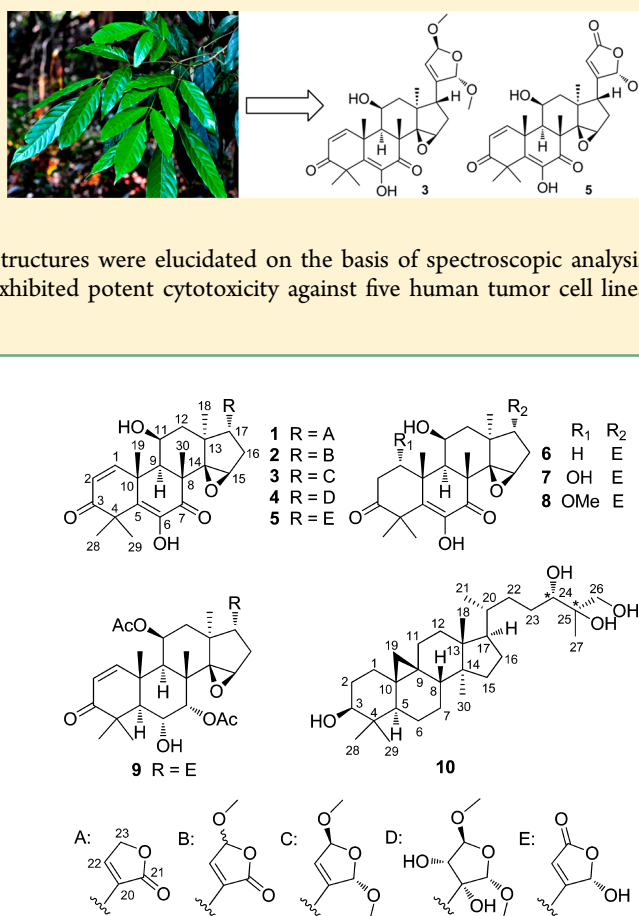
**ABSTRACT:** Nine new cedrelone limonoids, namely, walsuranolide B (1), 11 $\beta$ -hydroxy-23-O-methylwalsuranolide (2), yunnanolide A (3), 11 $\beta$ -hydroxyisowalsuranolide (5), 11 $\beta$ -hydroxy-1,2-dihydroisowalsuranolide (6), 1 $\alpha$ ,11 $\beta$ -dihydroxy-1,2-dihydroisowalsuranolide (7), 11 $\beta$ -hydroxy-1 $\alpha$ -methoxy-1,2-dihydroisowalsuranolide (8), and yunnanolide B (9), together with a new cycloartane triterpenoid, (24S\*,25R\*)-cycloartane-3 $\beta$ ,24,25,26-tetrol (10), were isolated from the leaves and twigs of *Walsura yunnanensis*. Their structures were elucidated on the basis of spectroscopic analysis and by comparison with literature data. Compounds 3 and 5 exhibited potent cytotoxicity against five human tumor cell lines with IC<sub>50</sub> values in the range 2.2–4.2  $\mu$ M.

Meliaceous limonoids have attracted interest due to their diverse structures and biological activities, such as insect antifeedant, antimalarial, cytotoxic, and 11 $\beta$ -HSD1 inhibitory effects.<sup>1,2</sup> The genus *Walsura* Roxb. (Meliaceae), comprising 30 to 40 species, is distributed primarily in mainland China, India, Malaysia, and Indonesia.<sup>3</sup> Chemical studies conducted in the past decade have shown that this genus is rich in biologically active limonoids. *Walsura yunnanensis* C. Y. Wu is endemic to the Xishuangbanna region of Yunnan Province, People's Republic of China.<sup>3</sup> Earlier investigations have shown the presence of limonoids and phenolic compounds.<sup>2a,4</sup> In a continuing study of the medicinal plants of Xishuangbanna for biologically active secondary metabolites, the leaves and twigs of *W. yunnanensis* were investigated. As a result, 10 new compounds (1–10) were isolated. This report describes the extraction, isolation, structure elucidation, and cytotoxicity evaluation of these compounds.

## RESULTS AND DISCUSSION

The leaves and twigs of *W. yunnanensis* were extracted three times with 95% EtOH. After removing the solvent, the ethanol extracts were suspended in water and then partitioned successively with petroleum ether, EtOAc, and *n*-BuOH. The EtOAc-soluble fraction was purified by column chromatography to produce nine new cedrelone limonoids (1–9) and one new cycloartane triterpenoid (10).

Compound 1 was obtained as colorless needles (CHCl<sub>3</sub>). Its HREIMS showed a molecular ion peak at *m/z* 454.1986 [M]<sup>+</sup> (calcd 454.1992), consistent with a molecular formula of C<sub>26</sub>H<sub>30</sub>O<sub>7</sub>, with 16 mass units more than that of the known compound 11 $\beta$ -hydroxycedrelone, isolated from the same



plant.<sup>2a</sup> The NMR data of 1 (Tables 1 and 2) were very close to those reported for 11 $\beta$ -hydroxycedrelone, with the only difference being the replacement of signals for the furan ring ( $\delta_{\text{H}}$  7.35 (d, *J* = 1.5 Hz), 7.15 and 6.15 s;  $\delta_{\text{C}}$  122.7, 139.5, 110.5, and 143.2) by those for an  $\alpha,\beta$ -unsaturated- $\gamma$ -lactone ring<sup>5</sup> [ $\delta_{\text{H}}$  4.71 (2H, m) and 7.13 (1H, s);  $\delta_{\text{C}}$  71.2, 133.3, 149.0, and 174.6] in 1. This conclusion was confirmed by the HMBC correlations between H-17/C-21 and H-23/C-21 as well as the ROESY correlations between H-11/H-9 and H-9/H<sub>3</sub>-18. Thus,

Received: November 23, 2013

Table 1.  $^1\text{H}$  NMR Spectroscopic Data of Compounds 1–5<sup>a</sup>

proton position	1 <sup>b</sup>	2 <sup>c</sup>	3 <sup>b</sup>	4 <sup>c</sup>	5 <sup>b</sup>
	(mult., <i>J</i> in Hz)	(mult., <i>J</i> in Hz)	(mult., <i>J</i> in Hz)	(mult., <i>J</i> in Hz)	(mult., <i>J</i> in Hz)
1	7.28, d (10.0)	7.30, d (10.0)	7.32, d (10.0)	7.24 <sup>f</sup>	7.32, d (10.0)
2	6.29, d (10.0)	6.30, d (10.0)	6.33, d (10.0)	6.30, d (9.8)	6.31, d (10.0)
9	2.85, s	2.82, s	2.85, s	3.02, s	2.86, s
11	5.04, d (6.3)	5.04, m	5.03, m <sup>f</sup>	5.02, m	5.12, d (6.0)
12 $\alpha$	2.26, dd (15.1, 6.3)	2.20, dd (15.2, 6.2)	2.36, dd (15.0, 5.8)	2.93, dd (15.0, 6.2)	2.34, dd (14.5, 6.0)
12 $\beta$	2.95, d (15.1)	2.91, d (15.2)	2.30, d (15.0)	2.57, d (15.0)	2.81, d (14.5)
15	4.08, s	4.08, s	4.08, s	4.20, s	4.11, s
16 $\alpha$	2.04, m	2.05, dd (13.3, 11.1)	1.95, dd (13.3, 11.6)	2.56, m	2.06, m
16 $\beta$	2.14, dd (13.5, 6.5)	2.15, dd (13.3, 6.4)	2.19, dd (13.3, 6.5)	2.25, dd (13.2, 6.2)	2.17, dd (13.4, 6.4)
17	3.02, m	2.98, m	2.64, dd (10.8, 6.5)	2.68, dd (10.7, 6.2)	3.26, dd (10.3, 6.4)
18	0.92, 3	0.90, s	1.05, s	1.70, s	1.01, s
19	1.81, s	1.81, s	1.80, s	1.79, s	1.82, s
21			5.30, s	5.06, s	6.43, s
22	7.13, s	7.01, s	5.73, s	4.65, dd (6.5, 4.4)	6.09, s
23	4.71, m <sup>d</sup>	5.93, s	5.77, s	5.45, d (4.4)	
28	1.88, s <sup>e</sup>	1.87, s	1.892, s	1.89, s	1.878, s
29	1.88, s <sup>e</sup>	1.88, s	1.888, s	1.88, s	1.884, s
30	1.78, s	1.78, s	1.75, s	1.77, s	1.78, s
OMe-21			3.40, s	3.39, s	
OMe-23		3.41, s	3.36, s	3.43, s	

<sup>a</sup>Recorded in  $\text{C}_5\text{D}_5\text{N}$ . <sup>b</sup>Recorded at 500 MHz. <sup>c</sup>Recorded at 600 MHz. <sup>d</sup>Double protons. <sup>e</sup>Overlapped by other signal. <sup>f</sup>Overlapped by solvent signal.

the structure of **1** was elucidated as depicted, and this compound has been named walsuranolide B.

Compound **2** was assigned a molecular formula of  $\text{C}_{27}\text{H}_{32}\text{O}_8$  on the basis of its HREIMS, or 30 mass units greater than that of **1**. The NMR data of **2** differed from those of **1** (Tables 1 and 2) only in the absence of signals for the oxymethylene ( $\text{CH}_2$ -23;  $\delta_{\text{H}}$  4.71, 2H, m;  $\delta_{\text{C}}$  71.2) in the  $\alpha,\beta$ -unsaturated- $\gamma$ -lactone ring and the presence of signals for dioxymethine [ $\delta_{\text{H}}$  5.93 (1H, s);  $\delta_{\text{C}}$  103.2] and methoxy ( $\delta_{\text{H}}$  3.41, 3H, s;  $\delta_{\text{C}}$  56.4) groups. The methoxy group was revealed to be located at C-23 by the HMBC correlation between  $\text{OCH}_3$ -23 and C-23, although its relative configuration remains undetermined. Further 2D NMR spectroscopic data analysis confirmed this assignment and was used to establish the structure of **2** as the 11 $\beta$ -hydroxy-23-O-methyl derivative of walsuranolide, 11 $\beta$ -hydroxy-23-O-methyl-walsuranolide.

Compound **3** was obtained as colorless needles ( $\text{CHCl}_3$ ). Its molecular formula,  $\text{C}_{28}\text{H}_{36}\text{O}_8$ , was established by the HREIMS peak at  $m/z$  500.2413  $[\text{M}]^+$  (calcd 500.2410), or 16 mass units greater than that of compound **2**. Comparison of its  $^1\text{H}$  and  $^{13}\text{C}$  NMR data (Tables 1 and 2) suggested that **3** differs only in the replacement of the ester carbonyl ( $\delta_{\text{C}}$  171.6, C-21) in **2** by a methoxymethine ( $\delta_{\text{H}}$  5.30,  $\delta_{\text{C}}$  109.7, CH-21;  $\delta_{\text{H}}$  3.40,  $\delta_{\text{C}}$  55.6, OMe-21). The ROESY correlations between  $\text{H}_3$ -18/H-23 and H-21/H-17 indicated H-23 and H-21 to be  $\alpha$ - and  $\beta$ -oriented, respectively. Therefore, the structure of compound **3** was established as shown, and this compound has been named yunnanolide A.

The molecular formula of compound **4** was determined to be  $\text{C}_{28}\text{H}_{38}\text{O}_{10}$  from its molecular ion peak at  $m/z$  534.2467  $[\text{M}]^+$  (calcd 534.2465) in the HREIMS, 34 mass units more than that of **3**. The 1D NMR data of **4** resembled closely those of **3** (Tables 1 and 2), except for the absence of signals for a  $\Delta^{20,22}$  double bond and the presence of signals for an oxymethine ( $\delta_{\text{H}}$  4.65, dd,  $J$  = 6.5, 4.4 Hz;  $\delta_{\text{C}}$  79.1; CH-22) and an oxygenated quaternary carbon ( $\delta_{\text{C}}$  82.3, C-20). These observations suggested **4** to be the 20,22-dihydroxy-20,22-dihydro derivative

of **3**. The  $^1\text{H}$ – $^1\text{H}$  COSY correlation between H-22 and H-23 as well as the HMBC correlations between  $\text{H}_2$ -16/C-20, H-21/C-17, H-21/C-23,  $\text{OCH}_3$ -21/C-21, and  $\text{OCH}_3$ -23/C-23 (Figure S25, Supporting Information) confirmed this inference. The ROESY correlations between H-17/H-21 and H-17/H-22 indicated that H-21 and H-22 are  $\beta$ -oriented, while the absence of any ROESY correlation between H-23 and H-21 was consistent with the presence of a H-23 $\alpha$  substituent. The structure of **4** (yunnanol A) was thus established as shown.

Compound **5** was obtained as a crystalline solid ( $\text{CHCl}_3$ ). Its molecular formula,  $\text{C}_{26}\text{H}_{30}\text{O}_8$ , was established by the HREIMS peak at  $m/z$  470.1951  $[\text{M}]^+$  (calcd 470.1941), showing 16 mass units more than that of isowalsuranolide isolated previously from this plant.<sup>2a</sup> The  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of **5** (Tables 1 and 2) were very similar to those of isowalsuranolide, with a major difference being the replacement of signals for a methylene ( $\delta_{\text{C}}$  19.6) in isowalsuranolide by those for an oxymethine ( $\delta_{\text{H}}$  5.12, d,  $J$  = 6.0 Hz;  $\delta_{\text{C}}$  66.4). These observations suggested that **5** is the 11-hydroxy derivative of isowalsuranolide, and the doublet for H-11 and a singlet for H-9 indicated a cis relationship between H-9 and H-11.<sup>2a</sup> Thus, the structure of **5** was deduced as 11 $\beta$ -hydroxyisowalsuranolide.

Compound **6** was obtained as colorless needles ( $\text{CHCl}_3$ ). Its HREIMS revealed a molecular ion peak at  $m/z$  472.2081  $[\text{M}]^+$  (calcd 472.2097), consistent with a molecular formula of  $\text{C}_{26}\text{H}_{32}\text{O}_8$ , having two mass units more than that of compound **5**. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of **6** (Tables 2 and 3) were very similar to those of **5**. Differences between the two compounds were the absence of signals for the  $\Delta^{1,2}$  double bond and the presence of signals for two additional  $\text{sp}^3$  methylenes ( $\delta_{\text{C}}$  36.2 and 33.6) in **6**. This compound was therefore assigned as a 1,2-dihydro derivative of **5**. Further 2D NMR spectroscopic data analysis confirmed the structure proposed for **6** as 11 $\beta$ -hydroxy-1,2-dihydroisowalsuranolide.

Compound **7** was obtained as colorless needles ( $\text{CHCl}_3$ ), and its molecular formula,  $\text{C}_{26}\text{H}_{32}\text{O}_9$ , was established from the HREIMS peak at  $m/z$  488.2041  $[\text{M}]^+$  (calcd 488.2046),

Table 2.  $^{13}\text{C}$  NMR Spectroscopic Data of Compounds 1–9

position	1 <sup>a</sup>	2 <sup>b</sup>	3 <sup>a</sup>	4 <sup>b</sup>	5 <sup>a</sup>	6 <sup>a</sup>	7 <sup>c</sup>	8 <sup>a</sup>	9 <sup>d</sup>
1	154.1	154.1	154.1	154.3	154.0	36.2	71.1	81.2	158.2
2	127.5	127.5	127.6	127.4	127.6	33.6	45.0	38.8	125.3
3	204.4	204.4	204.4	204.5	204.4	214.9	214.9	214.1	203.5
4	49.5	49.4	49.5	49.4	49.5	48.8 <sup>e</sup>	49.0	48.7	41.0
5	134.1	134.0	134.2	133.8	134.0	140.0	138.4	135.6	48.6
6	144.0	144.0	144.0	144.0	144.1	142.9	145.3	145.1	69.8
7	199.1	199.0	199.0	199.1	198.9	199.5	199.6	199.3	73.7
8	47.5	47.4	47.5	47.5	47.4	47.0	46.8	46.67	44.3
9	46.0	45.8	46.1	45.6	46.1	48.8 <sup>e</sup>	40.4	40.2	41.9
10	41.66	41.63	41.7	41.6	41.6	40.1	45.5	44.9	41.7
11	66.5	66.4	66.9	67.0	66.4	66.1	66.3	66.3	63.6
12	47.2	47.0	47.6	47.4	46.5	46.7	46.5	46.72	42.8
13	41.68	41.60	41.8	42.0	42.0	42.1	42.2	42.2	41.1
14	69.6	69.6	69.8	69.5	69.4	69.7	70.0	69.8	71.5
15	56.5	56.3	56.6	56.9	56.2	56.5	56.6	56.4	56.0
16	31.8	31.7	32.5	29.0	31.1	31.2	31.1	31.1	30.7
17	43.3	43.3	45.2	48.9	45.4	45.4	45.4	45.3	42.1
18	22.9	22.9	22.9	23.2	23.0	23.2	22.9	23.0	20.9
19	26.7	26.7	26.7	26.6	26.7	17.9	18.1	17.7	24.0
20	133.3	138.1	144.9	82.3	170.7	170.8	170.8	170.9	169.9
21	174.6	171.6	109.7	111.0	100.3	100.4	100.3	100.4	99.0
22	149.0	146.5	127.7	79.1	120.0	120.0	119.9	119.9	119.4
23	71.2	103.2	107.3	112.6	171.5	171.6	171.6	171.7	170.6
28	27.6	27.6	27.7	27.6	27.6	25.0	25.0	24.6	31.4
29	22.0	22.0	22.0	21.8	22.0	21.5	22.1	21.9	20.1
30	23.6	23.6	23.5	23.8	23.6	24.0	24.1	24.1	21.4
OMe-1								57.2	
OAc-7									170.1
									20.9
OAc-11									169.6
									21.0
OMe-21			55.6	55.2					
OMe-23		56.4	53.2	56.3					

<sup>a</sup>Recorded in  $\text{C}_5\text{D}_5\text{N}$  at 125 Hz. <sup>b</sup>Recorded in  $\text{C}_5\text{D}_5\text{N}$  at 150 Hz. <sup>c</sup>Recorded in  $\text{C}_5\text{D}_5\text{N}$  at 100 Hz. <sup>d</sup>Recorded in  $\text{DMSO}-d_6$  at 100 Hz. <sup>e</sup>Overlapped by other signal.

showing 16 mass units more than that of compound 6 (Tables 2 and 3). The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopic data observed for 7 were similar to those of compound 6. However, signals for  $\text{CH}_2$ -1 ( $\delta_{\text{H}}$  2.35 and 1.89;  $\delta_{\text{C}}$  36.2) and  $\text{CH}_2$ -2 ( $\delta_{\text{H}}$  2.85 and 2.69;  $\delta_{\text{C}}$  33.6) were replaced by proton signals at  $\delta_{\text{H}}$  4.68, 3.21, and 3.14 and carbon signals at  $\delta_{\text{C}}$  71.1 and 45.0, which suggested that 7 is either a 1- or 2-hydroxy derivative of 6. The HMBC correlations H-1/C-3, H-9/C-1, and  $\text{H}_3$ -19/C-1 indicated the hydroxy group is at C-1, and the ROESY correlation H-1/ $\text{H}_3$ -19 suggested that the hydroxy group is  $\alpha$ -oriented. Thus, 7 was determined as 1 $\alpha$ ,11 $\beta$ -dihydroxy-1,2-dihydroisowalsuranolide.

Compound 8 was obtained as colorless needles ( $\text{CHCl}_3$ ), and its HREIMS displayed a molecular ion peak at  $m/z$  502.2205  $[\text{M}]^+$  (calcd 502.2203), which corresponded to a molecular formula of  $\text{C}_{27}\text{H}_{34}\text{O}_9$ , representing 14 mass units more than that of compound 7. The 1D NMR spectroscopic data of 8 (Tables 2 and 3) were almost identical to those of compound 7, with the only difference being the presence of an additional methoxy group. The downfield shift of signals for the oxymethine ( $\delta_{\text{C}}$  C-1, from 71.1 to 81.2) suggested that this methoxy group is located at C-1, which was further supported by the HMBC correlations from  $\text{OCH}_3$ -1 ( $\delta_{\text{H}}$  3.34, s) to C-1 ( $\delta_{\text{C}}$  81.2) and H-1 ( $\delta_{\text{H}}$  3.83, d,  $J = 6.1$  Hz) to  $\text{OCH}_3$ -1 ( $\delta_{\text{C}}$

57.2). The ROESY correlation H-1/ $\text{H}_3$ -19 suggested that the methoxy group is  $\alpha$ -oriented. Compound 8 was therefore established as 11 $\beta$ -hydroxy-1 $\alpha$ -methoxy-1,2-dihydroisowalsuranolide.

Compound 9 was obtained as a white, amorphous powder, and its HREIMS displayed a molecular ion peak at  $m/z$  558.2461  $[\text{M}]^+$  (calcd 558.2465), corresponding to the molecular formula  $\text{C}_{30}\text{H}_{38}\text{O}_{10}$ . Comparison of the NMR spectroscopic data of 9 (Tables 2 and 3) with those of 5 showed that signals for the  $\alpha$ -hydroxy- $\alpha,\beta$ -unsaturated ketone group were absent, whereas those for three methines ( $\delta_{\text{H}}$  5.30, dd,  $J = 12.4, 2.5$  Hz; 4.775, m; 4.770, m;  $\delta_{\text{C}}$  69.8, 73.7, 63.6) and two acetoxy groups ( $\delta_{\text{H}}$  2.10, s; 1.94, s;  $\delta_{\text{C}}$  170.1, 20.9; 169.6, 21.0) were observed. These observations suggested 9 to be a 5,6,7-trihydrodiacetate derivative of 5. Further 2D NMR spectroscopic data analysis (Figures S59–S62, Supporting Information) not only confirmed this assignment but allowed the placement of the two acetoxy groups at C-7 and C-11, respectively, via HMBC correlations of H-7/ $\text{CH}_3\text{CO}$ -7 ( $\delta_{\text{C}}$  170.1) and H-11/ $\text{CH}_3\text{CO}$ -11 ( $\delta_{\text{C}}$  169.6). The strong ROESY correlation between H-11/H-9 suggested H-11 to be  $\alpha$ -oriented, while the ROESY correlations of H-6/ $\text{H}_3$ -19 and H-7/ $\text{H}_3$ -30 indicated that H-6 and H-7 are  $\beta$ -oriented. Thus, 9

Table 3.  $^1\text{H}$  NMR Spectroscopic Data of Compounds 6–9

proton position	6 <sup>a</sup> (mult., $J$ in Hz)	7 <sup>b</sup> (mult., $J$ in Hz)	8 <sup>a</sup> (mult., $J$ in Hz)	9 <sup>c</sup> (mult., $J$ in Hz)
1 $\alpha$	1.89, m			7.55, d (10.2)
1 $\beta$	2.35, dd (13.2, 9.6)	4.68, m	3.83, d (6.1)	
2 $\alpha$	2.85, dd (18.8, 9.2)	3.14, m	3.02, d (18.9)	5.93, d (10.2)
2 $\beta$	2.69, dd (18.8, 9.6)	3.21, m	2.87, dd (18.9, 6.1)	
5				2.43, m
6				5.30, dd (12.4, 2.5)
7				4.775, m
9	2.56, s	3.74, br s	3.45, s	2.29, br s
11	4.92, m	5.05, m <sup>d</sup>	4.92, d (6.0)	4.770, m
12 $\alpha$	2.23, m	2.20, m	2.33, dd (14.5, 6.0)	2.38, m
12 $\beta$	2.75, m	2.70, m	2.80, d (14.5)	1.68, m
15	4.09, s	4.13, m	4.11, s	3.33, m
16 $\alpha$	2.07, m	2.06, m	2.06, dd (13.3, 10.0)	1.86, m
16 $\beta$	2.16, m	2.15, m	2.14, dd (13.3, 6.2)	1.91, m
17	3.25, m	3.25, m	3.27, dd (10.0, 6.2)	2.41, m
18	1.02, s	1.07, s	1.07, s	0.92, s
19	1.61, s	1.67, s	1.58, s	1.46, s
21	6.44, s	6.27, s	6.47, s	5.96, s
22	6.10, s	6.03, s	6.08, s	6.01, s
28	1.77, s	2.11, s	1.89, s <sup>e</sup>	1.16, s
29	1.85, s	1.95, s	1.89, s <sup>e</sup>	1.05, s
30	1.74, s	1.82, s	1.77, s	1.34, s
OMe-1			3.34, s	
OAc-7				2.10, s
OAc-11				1.94, s

<sup>a</sup>Recorded in  $\text{C}_5\text{D}_5\text{N}$  at 500 Hz. <sup>b</sup>Recorded in  $\text{C}_5\text{D}_5\text{N}$  at 400 Hz. <sup>c</sup>Recorded in  $\text{DMSO}-d_6$  at 400 Hz. <sup>d</sup>Overlapped by solvent signal. <sup>e</sup>Overlapped by other signal.

was established structurally as depicted and was named yunnanolide B.

Compound **10** was isolated as an amorphous powder, with its molecular formula determined to be  $\text{C}_{30}\text{H}_{52}\text{O}_4$  on the basis of the HREIMS data obtained. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra (Table 4) showed this compound to have five tertiary methyl groups ( $\delta_{\text{H}}$  0.90, 1.00, 1.66, 1.12, and 1.24), one secondary methyl group ( $\delta_{\text{H}}$  1.02), two cyclopropylmethylene protons ( $\delta_{\text{H}}$  0.31, d,  $J = 3.5$  Hz; 0.54, d,  $J = 3.5$  Hz), and two typical upfield quaternary carbons (C-9,  $\delta_{\text{C}}$  20.3; C-10,  $\delta_{\text{C}}$  26.9). These observations indicated that **10** is a cycloartane triterpenoid. Detailed comparison of its NMR data with those of (24S)-25-methoxycycloartane-3 $\beta$ ,24-diol<sup>6</sup> suggested compound **10** to be its 26-hydroxy analogue. This was supported by the HMBC correlations (Figure 1A) of  $\text{H}_2$ -26/C-24,  $\text{H}_3$ -27/C-26, and H-24/C-26. Observation of a strong ROESY correlation between  $\text{H}_3$ -27 and H-24 and the absence of any ROESY correlation between  $\text{H}_2$ -23 and H-26 in its preferred configuration suggested the relative stereochemistry as 24S\* and 25R\* (Figure 1B1 and B2). This inference was further supported by comparing its  $^1\text{H}$  NMR data (in  $\text{CDCl}_3$ ; Figure S71, Supporting Information) for H-24 and  $\text{H}_2$ -26 [ca.  $\delta_{\text{H}}$  3.82 (d,  $J = 11.1$  Hz, 1H), 3.46 (m, 2H)] with those of a triterpenoid triol 24S,25R

Table 4.  $^1\text{H}$  and  $^{13}\text{C}$  NMR Spectroscopic Data (500 and 125 MHz,  $\text{C}_5\text{D}_5\text{N}$ ) for Compound 10

position	$\delta_{\text{H}}$ (mult., $J$ in Hz)	$\delta_{\text{C}}$	position	$\delta_{\text{H}}$ (mult., $J$ in Hz)	$\delta_{\text{C}}$
1 $\alpha$	1.54, m	32.8	16 $\alpha$	1.97, m	28.9
1 $\beta$	1.25, m		16 $\beta$	1.68, m	
2 $\alpha$	2.00, m	28.9	17	1.00, s <sup>b</sup>	53.3
2 $\beta$	1.36, m		18	1.00, s <sup>b</sup>	18.8
3	3.55, dd (4.5, 11.5)	78.3	19a	0.54, d (3.5)	30.4
4		41.5	19b	0.31, d (3.5)	
5	1.32, m	47.8	20	1.57, m	36.8
6 $\alpha$	1.59, m	21.8	21	1.02 <sup>c</sup>	19.0
6 $\beta$	0.79, m		22a	1.82, m	34.5
7 $\alpha$	1.06, m	26.7	22b	1.73, m	
7 $\beta$	1.30, m		23a	1.99, m	31.6
8	1.49, dd (4.6, 12.3)	48.5	23b	1.89, m	
9		20.3	24	4.19, m	76.6
10		26.9	25		75.1
11 $\alpha$	2.02, m	27.0	26a	4.32, d (10.6)	69.5
11 $\beta$	1.14, m		26b	4.13, d (10.6)	
12	1.61, m <sup>a</sup>	33.6	27	1.66, s	20.6
13		45.7	28	1.24, s	26.6
14		49.4	29	1.12, s	15.3
15 $\alpha$	1.27, m	36.2	30	0.90, s	19.9
15 $\beta$	2.15, m				

<sup>a</sup>Double protons. <sup>b</sup>Overlapped by other signal. <sup>c</sup>Signal partially obscured.

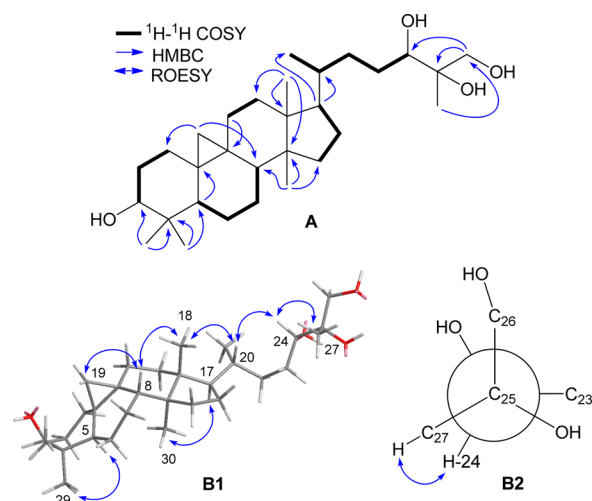


Figure 1. Selected  $^1\text{H}$ – $^1\text{H}$  COSY (bold bond), HMBC (A), and ROESY (B1 and B2) correlations of **10**.

(or 24R,25S) isomer [ca.  $\delta_{\text{H}}$  3.83 (d,  $J = 11.2$  Hz, 1H);  $\delta_{\text{H}}$  3.48 (m, 2H)] that possesses a similar ring D and is a side chain [for the 24S,25S (or 24R,25R) isomer]. However, the relevant signals for **10** appeared at ca.  $\delta_{\text{H}}$  3.65 (d,  $J = 11.2$  Hz, 1H), 3.49 (d,  $J = 11.2$  Hz, 1H), and  $\delta_{\text{H}}$  3.55 (1H).<sup>7</sup> Compound **10** was therefore determined to be (24S\*,25R\*)-cycloartane-3 $\beta$ ,24,25,26-tetrol.

Compounds **3**, **5**, **6**, **7**, **9**, and **10** were obtained in sufficient amounts to be evaluated for their cytotoxic activity against human myeloid leukemia (HL-60), hepatocellular carcinoma (SMMC-7721), lung cancer (A-549), breast cancer (MCF-7), colon cancer (SW480), and human bronchial epithelial (BEAS-2B) cell lines. Of the substances tested, only compounds **3** and



**5** exhibited potent cytotoxicity against the five human tumor cell lines, with  $IC_{50}$  values in the range 2.2–4.2  $\mu M$  (Table 5).

**Table 5. Cytotoxic Activity ( $IC_{50}$   $\mu M$ ) of Selected Isolated Compounds for Cancer Cell Lines**

compound	HL-60	SMMC-7721	A-549	MCF-7	SW480	BEAS-2B
<b>3</b>	3.6	2.4	3.7	4.2	3.5	5.0
<b>5</b>	3.1	2.2	2.6	3.9	2.4	9.4
cisplatin	1.9	5.3	8.7	>10	>10	>10

## EXPERIMENTAL SECTION

**General Experimental Procedures.** Melting points were obtained on an X-4 digital micromelting point apparatus and are uncorrected. Optical rotations were obtained with a JASCO P-1020 polarimeter. UV spectra were measured with a Shimadzu UV-2401A instrument. IR spectra (KBr) were determined on a Bruker Tensor-27 infrared spectrometer. 1D and 2D NMR spectra were recorded on Bruker AM-400, Bruker DRX-500, and Bruker Avance III 600 spectrometers with TMS as an internal standard. EIMS and HREIMS were recorded on an AutoSpec Premier P776 instrument, while ESIMS were measured with a Finnigan MAT 90 mass spectrometer. Semipreparative HPLC was carried out using a Waters system consisting of a 600 pump and a 2996 photodiode array detector. Silica gel (200–300 mesh, Qingdao Marine Chemical Factory, Qingdao, People's Republic of China), Sephadex LH-20 gel (40–70  $\mu m$ , Amersham Pharmacia Biotech AB, Uppsala, Sweden), and MCI gel (CHP20/P120, 75–150  $\mu m$ , high-porous polymer, Mitsubishi Chemical Corporation, Tokyo, Japan) were used for column chromatography.

**Plant Material.** The leaves and twigs of *W. yunnanensis* were collected from Xishuangbanna Tropical Botanical Garden (XTBG), Chinese Academy of Science (CAS), Mengla Country, Yunnan Province, People's Republic of China, in October 2012 and were identified by Chun-Fen Xiao (XTBG, CAS). A voucher specimen (no. 2012676) was deposited in the herbarium of XTBG.

**Extraction and Isolation.** The air-dried, powdered plant material (5.0 kg) was extracted three times (each for 7 days) with EtOH–H<sub>2</sub>O (95/5, v/v, 30 L) at room temperature. Removal of solvent of the combined extracts under a vacuum afforded a crude residue (300 g). The residue was then suspended in H<sub>2</sub>O (2 L) and partitioned with petroleum ether, EtOAc, and *n*-BuOH, successively. The EtOAc-soluble fraction (100 g) was subjected to silica gel column chromatography (CC) (MeOH–CHCl<sub>3</sub> from 0/1 to 1/1) to produce seven fractions (1–7). Fr. 2 (19.8 g) was applied to an MCI gel column (MeOH–H<sub>2</sub>O from 3/7 to 4/1) to yield five major fractions (2A–2E). Filtration of the precipitate from Fr. 2A (150 mg) yielded **10** (80 mg). Fr. 2C (500 mg) was chromatographed by Sephadex LH-20 CC, eluted with MeOH–H<sub>2</sub>O (from 2/3 to 4/1) to give Fr. 2C1 (15 mg), which was further purified by semipreparative HPLC (MeOH–H<sub>2</sub>O from 3/7 to 3/2) to yield **3** (6 mg). Fr. 3 (11.6 g) was separated over an MCI column eluted with MeOH–H<sub>2</sub>O (from 2/3 to 4/1) to give six major fractions (3A–3F). The precipitate from Fr. 3A (210 mg) yielded **9** (100 mg). Fr. 3B (1 g) was applied to a silica gel column eluted with petroleum ether–acetone (from 1/0 to 2/1) to give **5** (10 mg) and subfractions 3A1 (50 mg) and 3A2 (15 mg). Subfractions 3A1 and 3A2 were further purified by Sephadex LH-20 CC (MeOH–H<sub>2</sub>O from 3/7 to 3/2) and semipreparative HPLC (CH<sub>3</sub>CN–H<sub>2</sub>O from 2/3 to 7/3) to yield **2** (2 mg), **6** (8 mg), **7** (6 mg), and **8** (2 mg). Fr. 3C (20 mg) and 3D (55 mg) were purified initially by Sephadex LH-20 CC (MeOH–H<sub>2</sub>O from 1/4 to 1/1) and then by semipreparative HPLC (CH<sub>3</sub>CN–H<sub>2</sub>O from 1/4 to 13/7) to yield respectively **1** (6 mg) and **4** (3 mg).

**Walsuranolide B (1):** colorless needles (CHCl<sub>3</sub>); mp 128–130 °C;  $[\alpha]_D^{27}$  –3.7 (c 0.3, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 203 (4.20) nm; IR (KBr)  $\nu_{max}$  3426, 2935, 1753, 1679, 1628, 1451, 1352, 1251, 1122,

1092, 1054, 1035, 960, 756 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Tables 1 and 2; HREIMS  $m/z$  454.1986 [M]<sup>+</sup> (calcd for C<sub>26</sub>H<sub>30</sub>O<sub>7</sub>, 454.1992).

**11 $\beta$ -Hydroxy-23-O-methylwalsuranolide (2):** colorless needles (CHCl<sub>3</sub>); mp 115–117 °C;  $[\alpha]_D^{27}$  –2.1 (c 0.1, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 213 (4.01), 277 (3.84) nm; IR (KBr)  $\nu_{max}$  3442, 2924, 1767, 1631, 1456, 1383, 1260, 1095, 1035 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Tables 1 and 2; HREIMS  $m/z$  484.2105 [M]<sup>+</sup> (calcd for C<sub>27</sub>H<sub>32</sub>O<sub>8</sub>, 484.2097).

**Yunnanolide A (3):** colorless needles (CHCl<sub>3</sub>); mp 136–138 °C;  $[\alpha]_D^{27}$  –32.1 (c 0.1, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 203 (4.05), 276 (3.13) nm; IR (KBr)  $\nu_{max}$  3428, 2925, 1632, 1454, 1379, 1102, 1035 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Tables 1 and 2; HREIMS  $m/z$  500.2413 [M]<sup>+</sup> (calcd for C<sub>28</sub>H<sub>36</sub>O<sub>8</sub>, 500.2410).

**Yunnanol A (4):** colorless needles (CHCl<sub>3</sub>); mp 168–170 °C;  $[\alpha]_D^{28}$  –24.5 (c 0.1, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 211 (3.89), 278 (3.80) nm; IR (KBr)  $\nu_{max}$  3424, 2931, 1679, 1384, 1124, 1034, 992 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Tables 1 and 2; HREIMS  $m/z$  534.2467 [M]<sup>+</sup> (calcd for C<sub>28</sub>H<sub>38</sub>O<sub>10</sub>, 534.2465).

**11 $\beta$ -Hydroxyisowalsuranolide (5):** crystalline solid (CHCl<sub>3</sub>); mp 126–128 °C;  $[\alpha]_D^{27}$  –35.5 (c 0.1, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 220 (4.23), 278 (3.95) nm; IR (KBr)  $\nu_{max}$  3426, 2935, 1759, 1678, 1355, 1254, 1122, 1035, 950, 908 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Tables 1 and 2; HREIMS  $m/z$  470.1951 [M]<sup>+</sup> (calcd for C<sub>26</sub>H<sub>30</sub>O<sub>8</sub>, 470.1941).

**11 $\beta$ -Hydroxy-1,2-dihydroisowalsuranolide (6):** colorless needles (CHCl<sub>3</sub>); mp 130–132 °C;  $[\alpha]_D^{26}$  –55.7 (c 0.1, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 209 (4.08), 278 (3.86) nm; IR (KBr)  $\nu_{max}$  3431, 2925, 1746, 1703, 1679, 1631, 1384, 1253, 1135, 1034 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Tables 2 and 3; HREIMS  $m/z$  472.2081 [M]<sup>+</sup> (calcd for C<sub>26</sub>H<sub>32</sub>O<sub>8</sub>, 472.2097).

**1 $\alpha$ ,11 $\beta$ -Dihydroxy-1,2-dihydroisowalsuranolide (7):** colorless needles (CHCl<sub>3</sub>); mp 120–122 °C;  $[\alpha]_D^{27}$  –39.1 (c 0.1, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 214 (3.59), 279 (3.43) nm; IR (KBr)  $\nu_{max}$  3441, 2922, 2852, 1632, 1384 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Tables 2 and 3; HREIMS  $m/z$  488.2041 [M]<sup>+</sup> (calcd for C<sub>26</sub>H<sub>32</sub>O<sub>9</sub>, 488.2046).

**11 $\beta$ -Hydroxy-1 $\alpha$ -methoxy-1,2-dihydroisowalsuranolide (8):** colorless needles (CHCl<sub>3</sub>); mp 135–137 °C;  $[\alpha]_D^{27}$  –11.0 (c 0.1, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 210 (4.11), 279 (3.94) nm; IR (KBr)  $\nu_{max}$  3424, 2927, 1761, 1630, 1384, 1092, 1034 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Tables 2 and 3; HREIMS  $m/z$  502.2205 [M]<sup>+</sup> (calcd for C<sub>27</sub>H<sub>34</sub>O<sub>9</sub>, 502.2203).

**Yunnanolide B (9):** white, amorphous powder;  $[\alpha]_D^{26}$  +105.6 (c 0.1, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 221 (4.47) nm; IR (KBr)  $\nu_{max}$  3431, 2921, 1767, 1738, 1648, 1379, 1256, 1226, 1120, 1045 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Tables 2 and 3; HREIMS  $m/z$  558.2461 [M]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>38</sub>O<sub>10</sub>, 558.2465).

**(24S\*,25R\*)-Cycloartane-3 $\beta$ ,24,25,26-tetrol (10):** white, amorphous powder;  $[\alpha]_D^{26}$  +35.3 (c 0.1, MeOH); IR (KBr)  $\nu_{max}$  3430, 2966, 2936, 2866, 1635, 1449, 1377, 1337, 1287, 1121, 1081, 1043, 1021, 995, 916, 879 cm<sup>–1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data (C<sub>5</sub>D<sub>5</sub>N), see Table 4; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)  $\delta$  3.82 (1H, d,  $J$  = 11.1 Hz, H-26a), 3.47 (1H, m, H-24), 3.46 (1H, d,  $J$  = 11.1 Hz, H-26b), 3.27 (1H, m, H-3), 1.99 (2H, m, H-11), 1.09 (3H, s, H<sub>3</sub>-27), 0.963 (3H, s, H<sub>3</sub>-18), 0.96 (3H, s, H<sub>3</sub>-28), 0.89 (3H, s, H<sub>3</sub>-30), 0.88 (3H, d,  $J$  = 6.6 Hz, H<sub>3</sub>-21), 0.80 (3H, s, H<sub>3</sub>-29), 0.55 (1H, d,  $J$  = 3.9 Hz, H-19a), 0.32 (1H, d,  $J$  = 4.1 Hz, H-19b); HREIMS  $m/z$  476.3867 [M]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>52</sub>O<sub>4</sub>, 476.3866).

**Cytotoxicity Assays.** The MTS method was used for assessing the cytotoxicity of the compounds against five tumor cell lines (HL-60 human myeloid leukemia, SMMC-7721 hepatocellular carcinoma, A-549 lung cancer, MCF-7 breast cancer, and SW480 colon cancer). All cells were cultured in RPMI 1640 medium containing 10% fetal bovine serum and 100 U/mL penicillin/streptomycin in a humidified incubator in a 5% CO<sub>2</sub> atmosphere at 37 °C. Then, 100  $\mu L$  of adherent cells was seeded into each well (1  $\times$  10<sup>4</sup> cells/well) of 96-well cell culture plates and allowed to adhere for 12 h before test drug addition. Each tumor cell line was exposed to a test compound at concentrations of 0.064, 0.32, 1.6, 8, and 40  $\mu M$  in DMSO in triplicate for 48 h, with cisplatin as the positive control. After 48 h incubation, 20  $\mu L$  of MTS [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxy-

phenyl)-2-(4-sulfophenyl)-2H-tetrazolium] solution was added to each well, which were incubated for another 4 h to give a formazan product. Then 100  $\mu$ L of 20% SDS was added to each well and incubated 12 h at room temperature for the formazan product to dissolve completely. The OD value of each well was measured at 490 nm using a Biorad 680 instrument. The  $IC_{50}$  value of each compound was calculated by the Reed and Muench method.<sup>8</sup>

## ■ ASSOCIATED CONTENT

### ■ Supporting Information

This material (HREIMS, <sup>1</sup>H and <sup>13</sup>C NMR, and 2D NMR spectra of limonoids (**1–9**) and (24S\*,25R\*)-cycloartane-3 $\beta$ ,24,25,26-tetrol (**10**)) is available free of charge via the Internet at <http://pubs.acs.org>.

## ■ AUTHOR INFORMATION

### Corresponding Authors

\*(S.-G. Liao) Tel: +86-851-6908468. Fax: +86-851-6908468-0. E-mail: [lishangg@163.com](mailto:lishangg@163.com).

\*(Y.-K. Xu) Tel: +86-691-8713169. Fax: +86-691-871-3061. E-mail: [xyk@xtbg.ac.cn](mailto:xyk@xtbg.ac.cn).

### Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

CAS 135 program (XTBG-F02), NSFC-U1302222, and Ethnobotanical Investigation of Plants for Industrialization in Southwest China (SQ2012FY4910027) are gratefully acknowledged.

## ■ REFERENCES

- (1) Tan, Q. G.; Luo, X. D. *Chem. Rev.* **2011**, *111*, 7437–7522.
- (2) (a) Luo, X. D.; Wu, S. H.; Ma, Y. B.; Wu, D. G. *J. Nat. Prod.* **2000**, *63*, 947–951. (b) Yin, S.; Wang, X. N.; Fan, C. Q.; Liao, S. G.; Yue, J. M. *Org. Lett.* **2007**, *9*, 2353–2356. (c) Jiang, J. H.; Pan, Y. Q.; Ma, Y. J.; Chen, Y. G. *J. Anhui Agric. Sci.* **2008**, *36*, 14142–14143. (d) Zhou, Z. W.; Yin, S.; Zhang, H. Y.; Fu, Y.; Yang, S. P.; Wang, X. N.; Wu, Y.; Tang, X. C.; Yue, J. M. *Org. Lett.* **2008**, *10*, 465–458. (e) Awang, K.; Yusoff, M.; Mohamad, K.; Chong, S. L.; Ng, S. W. *Acta Crystallogr. E* **2009**, *65*, O1166–U3323. (f) Rao, M. S. A.; Suresh, G.; Yadav, P. A.; Prasad, K. R.; Nayak, V. L.; Ramakrishna, S.; Rao, C. V.; Babu, K. S. *Tetrahedron Lett.* **2012**, *53*, 6241–6244. (g) Sichaem, J.; Aree, T.; Khumkratok, S.; Jong-aramruang, J.; Tip-pyang, S. *Phytochem. Lett.* **2012**, *5*, 665–667. (h) Han, M. L.; Zhang, H.; Yang, S. P.; Yue, J. M. *Org. Lett.* **2012**, *14*, 486–489. (i) Nugroho, A. E.; Okuda, M.; Yamamoto, Y.; Hirasawa, Y.; Wong, C. P.; Kaneda, T.; Shiota, O.; Hadi, A. H. A.; Morita, H. *Tetrahedron* **2013**, *69*, 4139–4145. (j) Han, M. L.; Shen, Y.; Wang, G. C.; Leng, Y.; Zhang, H.; Yue, J. M. *J. Nat. Prod.* **2013**, *76*, 1319–1327.
- (3) Chen, S. K.; Li, H.; Chen, B. Y. In *Zhongguo Zhiwu Zhi*; Science Press: Beijing, 1997; Vol. 43 (3), p 62.
- (4) Jiang, L. H. *Chem. Nat. Compd.* **2013**, *48*, 1013–1016.
- (5) Yang, M. H.; Wang, J. S.; Luo, J. G.; Wang, X. B.; Kong, L. Y. *J. Nat. Prod.* **2009**, *72*, 2014–2018.
- (6) Ukiya, M.; Akihisa, T.; Yasukawa, K.; Kasahara, Y.; Kimura, Y.; Koike, K.; Nikaido, T.; Takido, M. *J. Agric. Food Chem.* **2001**, *49*, 3187–3197.
- (7) Kennedy, E. M.; P'Pool, S. J.; Jiang, J. H.; Sliva, D.; Minto, R. E. *J. Nat. Prod.* **2011**, *74*, 2332–2337.
- (8) Reed, L. J.; Muench, H. *Am. J. Hyg.* **1938**, *27*, 493–497.